Gelatin

**Add the following:**

> Portions of this monograph that are national USP text, and are not part of the harmonized text, are marked with symbols (•) to specify this fact.

**DEFINITION**

Purified protein obtained from collagen of animals (including fish and poultry) by partial alkaline and/or acid hydrolysis, by enzymatic hydrolysis, or by thermal hydrolysis.

The hydrolysis leads to gelling or non-gelling grades. This monograph covers both the gelling and non-gelling grades as well as hydrolyzed gelatin.

**IDENTIFICATION**

- **A.**
  
  **Sample solution:** Dissolve 1.00 g in carbon dioxide-free water at about 55°, dilute with the same solvent to 100 mL, and keep at 55°. Save the unused portion of this solution for use in the test for pH.
  
  **Analysis:** To 2 mL of Sample solution add 0.05 mL of a 125-g/L solution of copper sulfate pentahydrate. Mix, and add 0.5 mL of an 85-g/L solution of sodium hydroxide.
  
  **Acceptance criteria:** A violet color is produced.

- **B.**
  
  **Sample:** 0.5 g
  
  **Analysis:** Place the Sample in a test tube of about 15 mm internal diameter and add 10 mL of water. Allow to stand for 10 min, heat at 60° for 13 min, and keep the tube upright at 0° for 6 h. Invert the tube.
  
  **Acceptance criteria:** The contents do not flow out immediately for gelling grades. The contents immediately flow out for non-gelling grades.

**Change to read:**

- **C. FOR NON-GELLING GRADES**
  
  **Medium:** pH 6.8 phosphate buffer. Mix 77.3 mL of a 71.5-g/L solution of disodium hydrogen phosphate dodecahydrate with 22.7 mL of a 21-g/L solution of citric acid.
  
  **Sample:** 0.5 g
  
  **Analysis:** Place the Sample in a 250-mL bottle. Add 10 mL of water and 5 mL of sulfuric acid. Place the bottle, partly but not completely closed (for example, using a watch glass), in an oven at 105° for 4 h. Allow to cool, and add 200 mL of water. Adjust to a pH of 6.0–8.0 using a 200-g/L solution of sodium hydroxide. Place 2 mL of the solution in a test tube, and add 2 mL of oxidizing reagent [14-g/L solution of chloramine-T (NHCl)] and 2 mL of color reagent (prepared immediately before use). Mix, and allow to stand for 20 min. Add 2 mL of color reagent, and slowly add 6.3 mL of 2-propanol. Mix, and place in a water bath at 60° for about 15 min.

**SPECIFIC TESTS**

- **pH (791)**
  
  **Sample:** Use the Sample solution prepared in Identification test A.
  
  **Acceptance criteria:** 3.8–7.6 at 55°

- **WATER CONDUCTIVITY (645)**
  
  **Sample:** 1.0% Solution at 30 ± 1.0°
  
  **Analysis:** Determine without the use of the temperature compensation.
  
  **Acceptance criteria:** NMT 1 mS · cm⁻¹

**Change to read:**

- **SULFUR DIOXIDE**
  
  **Sample:** 25.0 g
  
  **Titrimetric system**
  
  **Mode:** Direct titration
  
  **Titrant:** 0.1 M sodium hydroxide VS bromophenol blue TS
  
  **Endpoint detection:** Visual
  
  **Analysis:** Introduce 150 mL of water into the flask (A) (see Figure 1), and pass carbon dioxide through the whole system for 15 min at a rate of 100 mL/min. To 10 mL of hydrogen peroxide solution [30 g/L of hydrogen peroxide (H₂O₂)] add 0.15 mL of bromophenol blue TS. Add 0.1 M sodium hydroxide until a viole-blue color is obtained, without exceeding the endpoint. Place the solution in the test tube (D). Without interrupting the stream of carbon dioxide, remove the funnel (B), and introduce through the opening into the flask (A) the Sample with the aid of 100 mL of water. Add through the funnel 80 mL of dilute hydrochloric acid [73 g/L of hydrogen chloride (HCl)], and boil for 1 h. Open the tap of the funnel, stop the flow of carbon dioxide, and also stop the heating and the cooling water. Transfer the contents of the test tube with the aid of a little water to a 200-mL wide-necked, conical flask. Heat on a water bath for 15 min, and allow to cool. Add 0.1 mL of bromophenol blue TS, and titrate with Titrant until the color changes from yellow to violet-blue.

**Figure 1. Apparatus for determination of sulfur dioxide.**

Carry out a blank titration, and calculate the content of sulfur dioxide, in ppm:

\[
\text{Result} = \left( V_2 - V_1 \right) \times \left( m/W \right) \times 32,030
\]

\[ V_1 = \text{volume of 0.1 M sodium hydroxide consumed by the Sample (mL)} \]
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Change to read:

- **PEROXIDES**
  Peroxidase transfers oxygen from peroxides to an organic redox indicator which is converted to a blue oxidation product. The intensity of the color obtained is proportional to the quantity of peroxide, and can be compared with a color scale provided with the test strips to determine the peroxide concentration.

  **Peroxide test strips:** Use commercial test strips with a suitable scale covering the range of 0–25 ppm of peroxide.

  **Sample:** 20.0 ± 0.1 g

  **Suitability test:** Dip a test strip for 1 s into hydrogen peroxide standard solution \( [(2,5)-][\text{ppm} \text{ of peroxide (H}_2\text{O}_2)] \ (\text{prepared by dilution of hydrogen peroxide (H}_2\text{O}_2)) \text{, such that the reaction zone is properly wetted. Remove the test strip, shake off excess liquid, and after 15 s compare the reaction zone with the color scale provided. The test strips are suitable if the color matches that of the 2,5 ppm concentration.}

  **Analysis:** Weigh the sample in a beaker, and add 80.0 ± 0.2 mL of water. Stir to moisten all the gelatin, and allow the sample to stand at room temperature for 1–3 h. Cover the beaker with a watch glass. If the sample is not dissolved completely, place the beaker in a water bath at 65 ± 2° for 20 ± 5 min to dissolve the sample. Stir the contents of the beaker with a glass rod to achieve a homogeneous solution. Dip a test strip for 1 s into the test solution, such that the reaction zone is properly wetted. Remove the test strip, shake off excess liquid, and after 15 s compare the reaction zone with the color scale provided. Multiply the concentration read from the color scale by a factor of 5 to calculate the concentration, in ppm, of peroxide in the test substance.

  **Acceptance criteria:** NMT 10 ppm

- **GEL STRENGTH (BLOOM VALUE)**
  For gelling grades

  The gel strength is expressed as the mass in grams necessary to produce the force which, applied to a plunger 12.7 mm in diameter, makes a depression 4 mm deep in a gel with a concentration of 6.67% m/m and matured at 10°.

  **Sample:** 7.5 g

  **Apparatus:** Texture analyzer or gelometer with a cylindrical piston 12.7 ± 0.1 mm in diameter with a plane pressure surface and a sharp bottom edge, and a bottle 59 ± 1 mm in internal diameter and 85 mm high. Adjust the apparatus according to the manufacturer’s manual.

  **Settings**

  - **Distance:** 4 mm
  - **Test speed:** 0.5 mm/s

  **Analysis:** Place the sample in a bottle. Add 105 mL of water, place a watch glass over the bottle, and allow to stand for 1–4 h. Heat in a water bath at 65 ± 2° for 15 min. While heating, stir gently with a glass rod. Ensure that the solution is uniform and that any condensed water on the inner walls of the bottle is incorporated. Allow to cool at room temperature for 15 min, and transfer the bottle to a thermostatically controlled bath at 10.0 ± 0.1°, fitted with a device to ensure that the platform on which the bottle stands is perfectly horizontal. Close the bottle with a rubber stopper, and allow to stand for 17 ± 1 h. Remove the sample bottle from the bath, and quickly wipe the water from the exterior of the bottle. Center the bottle on the platform of the apparatus so that the plunger contacts the sample as nearly at its midpoint as possible, and start the measurement.

  **Acceptance criteria:** NMT 50 ppm

- **IRON**

  **Atomic absorption spectrometry, standard additions method**

  **Iron standard solution** (8 ppm): Dissolve 80 mg of iron in 50 mL of hydrochloric acid [220 g/L of hydrogen chloride (HCl)], and dilute with water to 1000.0 mL. Immediately before use, prepare a 1:10 dilution with water.

  **Sample solution:** In a conical flask add 10 mL of hydrochloric acid [37% m/m of hydrogen chloride (HCl)] to 5.00 g of the substance to be examined. Close the flask, and place in a water bath at 75°–80° for 2 h. (If necessary for proper solubilization, the gelatin may be allowed to swell after addition of the acid and before heating, the heating time may be prolonged, and a higher temperature may be used.) Allow to cool, and adjust the contents of the flask with water to 100.0 g.

  **Instrumental conditions**

  (See Spectrophotometry and Light-Scattering (851).)

  **Analytical wavelength:** 248.3 nm

  **Acceptance criteria:** NMT 30 ppm

Change to read:

- **CHROMIUM**

  **Atomic absorption spectrometry, standard additions method**

  **Chromium standard solution** (100 ppm): 0.283 mg/mL of potassium dichromate (K2Cr2O7) in water

  **Standard solutions:** Prepare the solutions using the Chromium standard solution, diluted with water if necessary.

  **Sample solution:** Use the sample solution described in the test for Iron.

  **Instrumental conditions**

  (See Spectrophotometry and Light-Scattering (851).)

  **Analytical wavelength:** 357.9 nm

  **Acceptance criteria:** NMT 10 ppm

- **ZINC**

  **Atomic absorption spectrometry, standard additions method**

  **Zinc standard solution** (10 ppm): Dissolve 0.440 g of zinc sulfate heptahydrate and 1 mL of acetic acid (300 g/L of C2H4O2) in water, and dilute to 100.0 mL. Immediately before use, prepare a 1:100 dilution in water.

  **Standard solutions:** Prepare the solutions using the Zinc standard solution, diluted if necessary with water.

  **Sample solution:** Use the sample solution described in the test for Iron.

  **Instrumental conditions**

  (See Spectrophotometry and Light-Scattering (851).)
Analytical wavelength: 213.9 nm
Acceptance criteria: NMT 30 ppm

- **Loss on Drying (731)**
  - **Sample:** 5.0 g
  - **Analysis:** Dry the sample in an oven at 105°C for 16 h.
  - **Acceptance criteria:** NMT 15%

- **Microbial Enumeration Tests (61) and Tests for Specified Microorganisms (62):** The total bacterial count does not exceed 10^3 cfu/g, the total yeasts and molds count does not exceed 10^2 cfu/g, and the tests for *Salmonella* species and *Escherichia coli* are negative.

**Additional Requirements**
- **Storage:** Protect from heat and moisture.
- **Labeling:** The label states the Gel Strength (Bloom Value) or that it is a non-gelling grade.