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ANALYTICAL TOOLBOX REQUIREMENTS

to enable the synthesis and release of large synthetic peptides

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A LEADING SPECIALIST FOR DRUG SUBSTANCES

- Contract development and manufacturing organization (CDMO)
- Broad capabilities in Peptides and Oligonucleotides (TIDES) as active pharmaceutical ingredients (API)
- Long-term partnerships with pharmaceutical and biotech companies
- Focused on chemical synthesis, committed to innovation
- Annual sales of CHF 577.3 million in 2023 and over 2,000 colleagues globally
- Reliable supply of APIs for WHO essential medicines benefitting patients worldwide



High quality GMP manufacturing

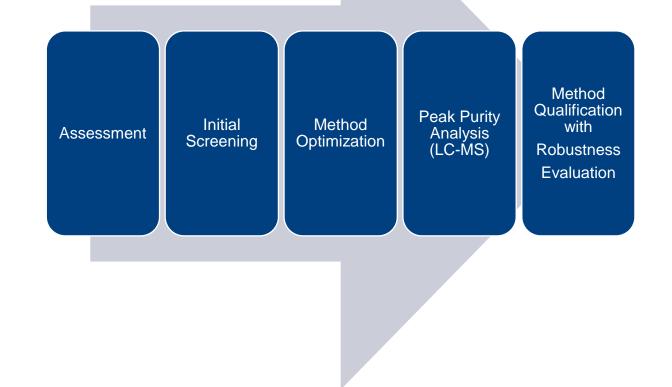




COMPLEX THERAPEUTIC PEPTIDES REQUIRE HIGHLY SELECTIVE PROCEDURES

- Critical quality attributes (CQAs):
- ➤ Purity
- Related impurities
- ➤ Assay
- Output:
- Identification of suitable procedure conditions
- Understanding effect of procedure parameters
- Initial analytical control strategy

Stage 1 (procedure design) of analytical life cycle <USP 1220>

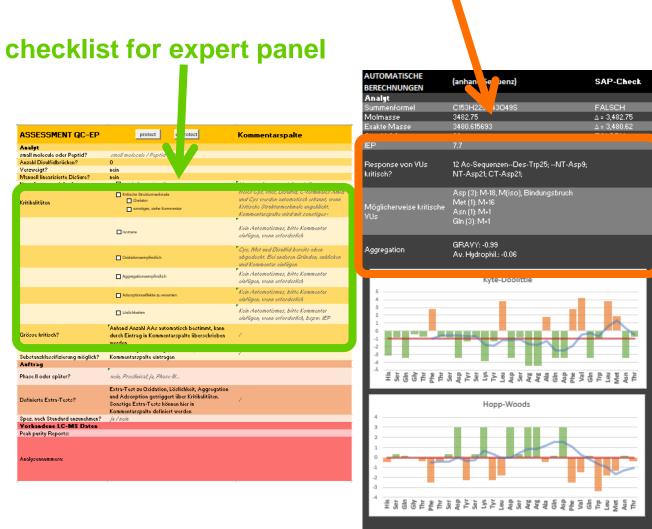




physico-chemical properties

ASSESSMENT FOR SYSTEMATIC USE OF PRIOR KNOWLEDGE

- In-silico predictions of physicochemical properties based on peptide sequence
- Expert panel evaluation based on molecular structure, synthesis route etc.
- Predict impurities
- Oxidation/aggregation potential
- Additional tests for procedure development
- Structural similarities with other APIs





DE NOVO METHOD DEVELOPMENT HAS MULTIPLE KEY OBJECTIVES

Initial Screening Method Optimization

01

03

Selectivity/Resolution limiting co-eluting impurities 04

High-Resolution MS-compatibility choice and conc. additives

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Sens S/N ≥

Sensitivity S/N ≥ 10 05 R

06

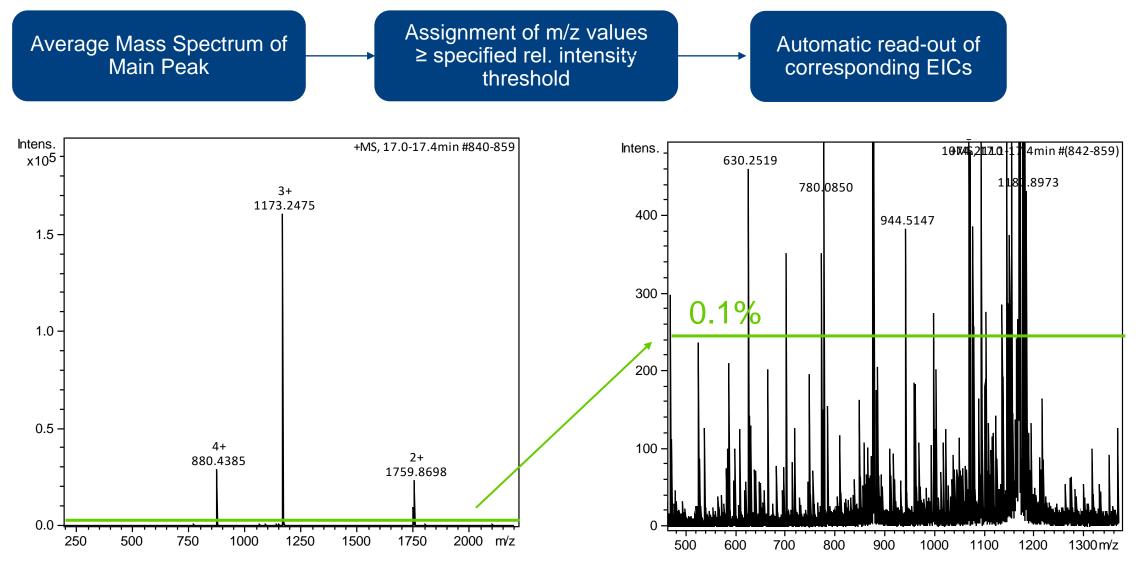
Robustness & Applicability Reproducibility over time

Stability indicating

detects related (degradation) impurities

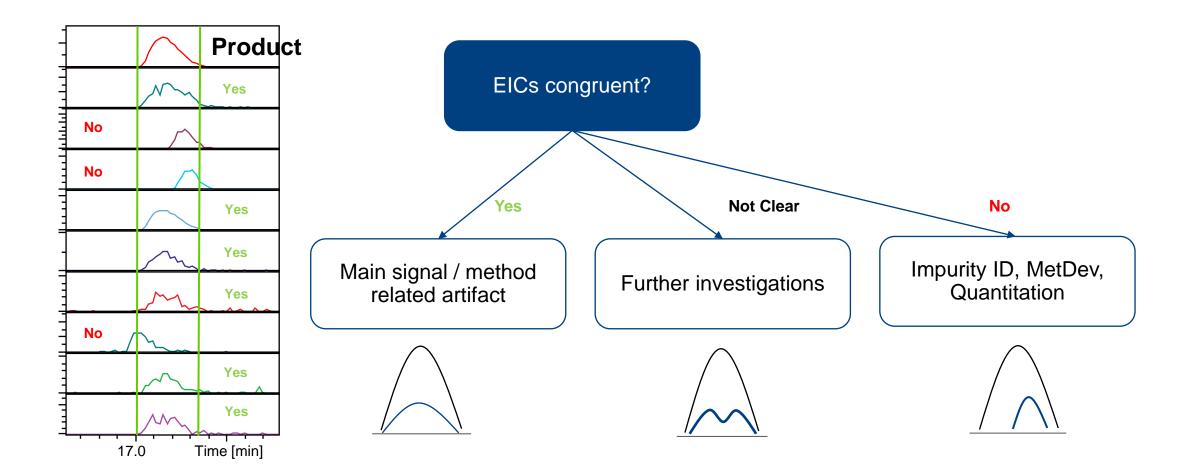
State of the Art Transferable to other labs

PEAK PURITY (LC-MS) FOR CO-ELUTING IMPURITIES



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PEAK PURITY (LC-MS) FOR CO-ELUTING IMPURITIES





INITIAL REQUIREMENT PURITY AND RELATED IMPURITIES

Co-elution of impurities with each other summed UV peaks may exceed specified limits ⇒ unfavorable

Co-elution of impurities with main peak

Impurity may exceed specified limit without detection!

⇒ unfavorable

A

Identification of impurities

Identity might explain origin and stability behaviour

⇒ important

EP-Monograph 2034 "Substances for Pharmaceutical Use"

Table 2034.-2. – Reporting, identification and qualification of organic impurities in peptides obtained by chemical synthesis

Reporting	Identification	Qualification
threshold	threshold	threshold
> 0.1 per cent	> 0.5 per cent	> 1.0 per cent

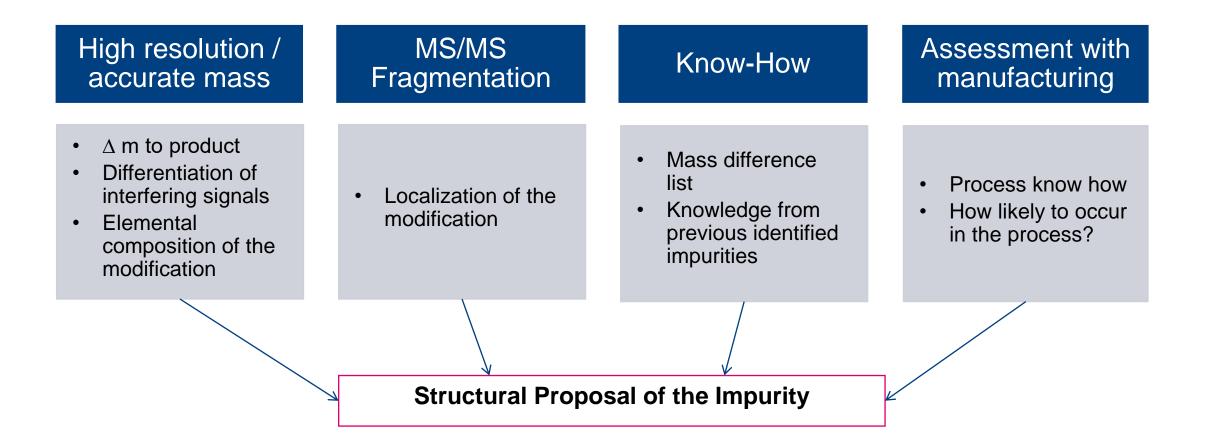
Purity: >95% (target)

Related Impurities:

- Specification for all impurities that may occur above 0.5%
- General limit for non-specified impurities < 0.5%
- Synthesis of impurity references

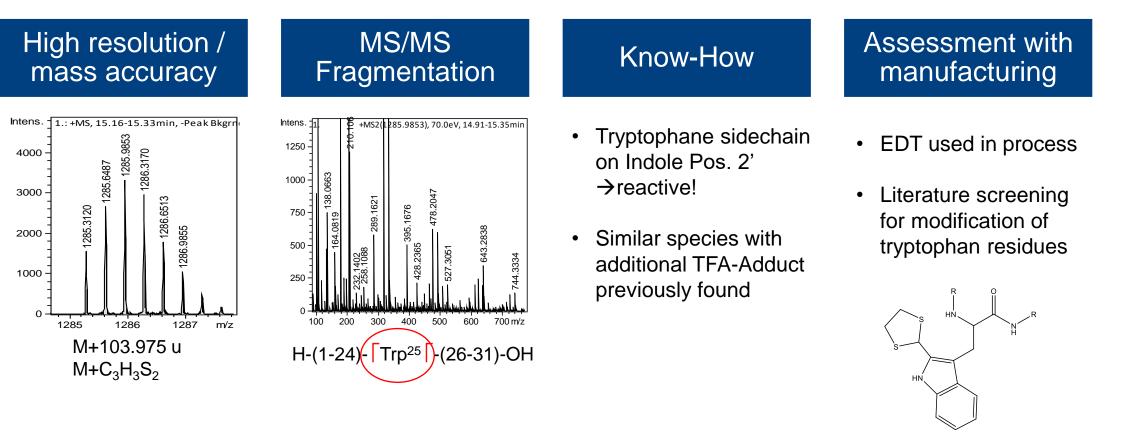


IDENTIFICATION BY LC-MS/MS – PROCEDURAL APPROACH





IDENTIFICATION BY LC-MS/MS EXAMPLE OF UNKNOWN IMPURITY M+104

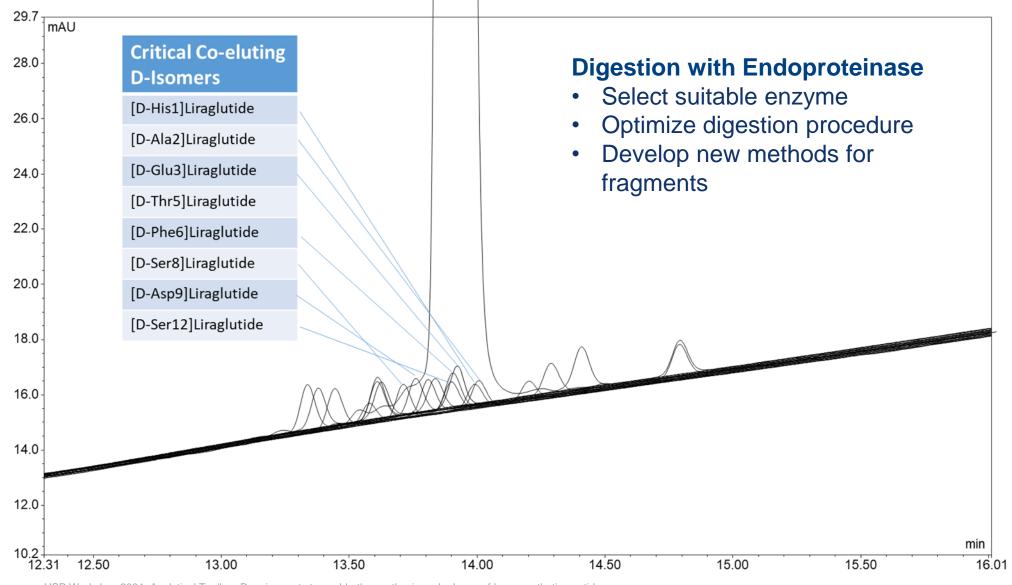


[2-(1,3-Dithiolan-2-yl)-Trp²⁵]-Liraglutide



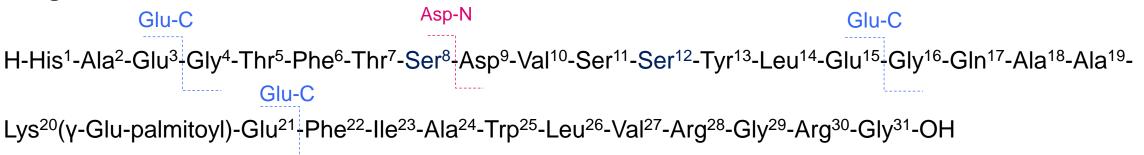
STEREOISOMERS POSE A CHALLENGE FOR LIRAGLUTIDE

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ENZYMATIC METHODS FOR D-ISOMERS

Liraglutide



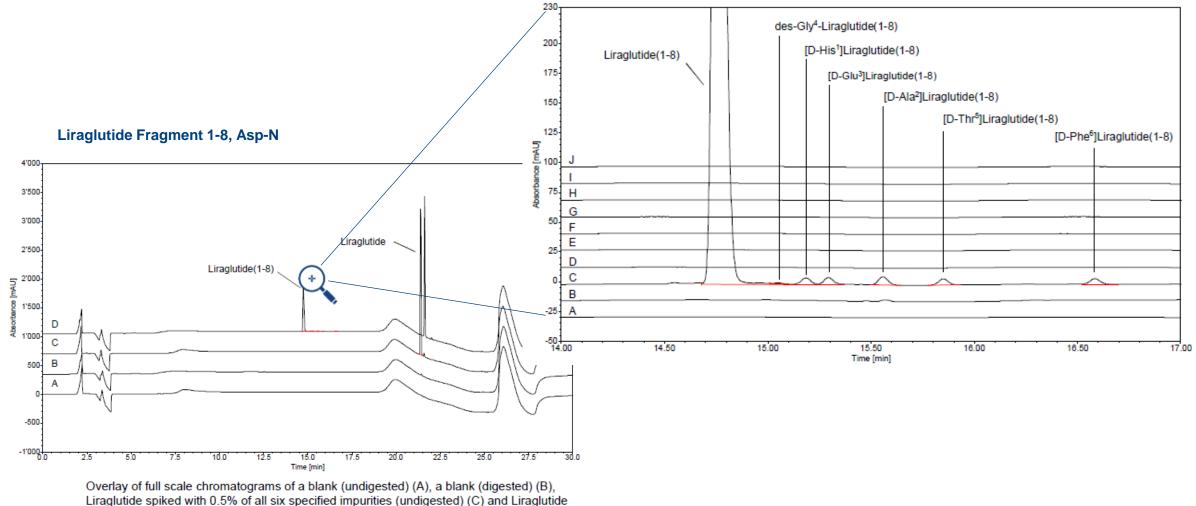
Asp-N (Fragment 1-8)	Glu-C (Fragment 4-15)	3 rd impurity method (w/o enzyme)
	[D-Ser8] Liraglutide	[D-Glu15] Liraglutide [D-Lys20] Liraglutide
[D-His1]Liraglutide [D-Ala2] Liraglutide	[D-Asp9] Liraglutide	[D-Glu21] Liraglutide
[D-Glu3] Liraglutide	[D-Ser12] Liraglutide	[D-Phe22] Liraglutide

[D-Thr5] Liraglutide

Aften continue of digest and development of HPLC methods, spiking experiments were conducted to identify the D-isomers fragments



HPLC METHOD FOR D-ISOMERS OF ASP-N DIGEST



spiked with 0.5% of all six specified impurities (digested) (D)



BATCH RESULTS WITH ENZYMATIC METHODS

		[D-His1]	[D-Ala2]	[D-Glu3]	[D-Thr5]	[D-Phe6]	[D-Ser8]	[D-Ser12]	[D-Glu15]	[D-Glu21]	[D-Lys20]	[D-Phe22]
	Batch	[%]	[%]	[%]	[%]	[%]	[%]	[%]	[%]	[%]	[%]	[%]
	Batch 9	0.13	<	<	<	<	0.27	0.06	<	<	<	0.07
	Batch 8	0.13	<	<	<	<	0.28	0.07	<	<	<	0.06
Ļ	Batch 7	0.13	<	<	<	<	0.27	0.07	<	<	<	<
[Batch 6	0.10	<	<	<	<	0.26	0.07	<	<	<	0.14
	Batch 5	0.11	<	<	<	<	0.27	0.06	<	<	<	0.14
Pr.Dev –	Batch 4	0.11	<	<	<	<	0.27	0.07	<	<	<	0.13
	Batch 3	0.12	<	0.05	<	<	0.30	0.08	<	0.05	<	0.16
	Batch 2	0.15	0.06	0.05	<	<	0.29	0.07	<	0.05	<	0.06
Ļ	Batch 1	0.19	0.07	0.07	<	<	0.30	0.08	0.06	<	<	0.07
	GS63P81	0.70	<	<	<	0.13	1.23	0.47	<	0.05	0.05	<
	JS69S28	0.47	<	0.41	<	0.09	1.20	0.45	<	<	<	<
RLD -	JS67Y95 (EU)	0.40	<	<	<	0.14	1.29	0.47	<	<	<	<
	JS68C66 (USA)	0.34	<	<	<	0.16	1.33	0.50	<	0.05	<	<
	JS68L68 (USA)	0.29	<	<	<	0.15	1.25	0.45	<	0.05	<	<
Ĺ	JS67T64 (USA)	0.37	<	<	<	0.15	1.36	0.50	<	0.06	<	<



ANALYTICAL TOOLBOX FOR PHASE III

Output: HPLC Purity Method Set of orthogonal methods to Determine purity of material determine purity, related Identify related impurities by MS 01 impurities & assay Quantify related impurities by LC UV Determine assay by HPLC **LC MS Quantitation** Co-eluting impurities unequivocally **Enzymatic Digest** 02 identified and quantified Impurities under main peak can be Reduce complexicity of large peptides quantified Resolve isomers that are challenging Necessary when selectivity can not to detect by LC-MS/MS 03 be achieved chromatographically Quantify impurities by LC-UV

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REQUIREMENTS FOR PHASE III

01

Purity Purity ≥ 99.0% target



Process Understanding Origin and fate of impurities better understood



Related Impurities Report impurities ≥ 0.05 % Identify impurities ≥ 0.10 %



06

Comprehensive

methods enable detection of any Impurity or degradation product



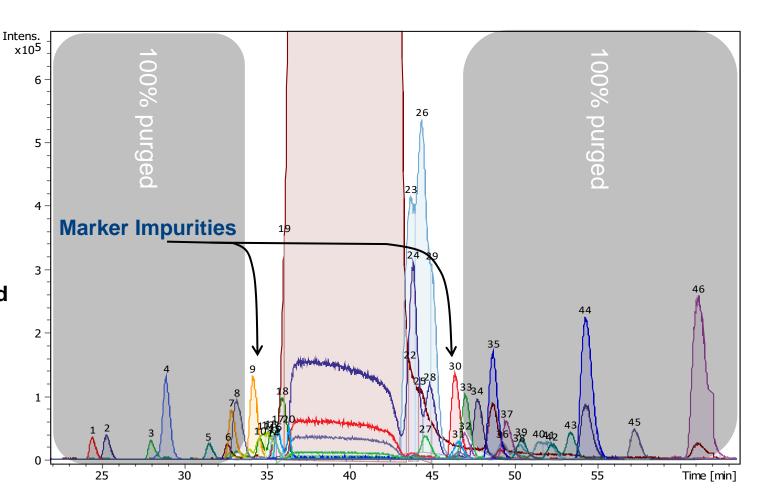
Specificity specified impurities synthesized **Robustness**

methods developed and optimized to ensure reproducible results



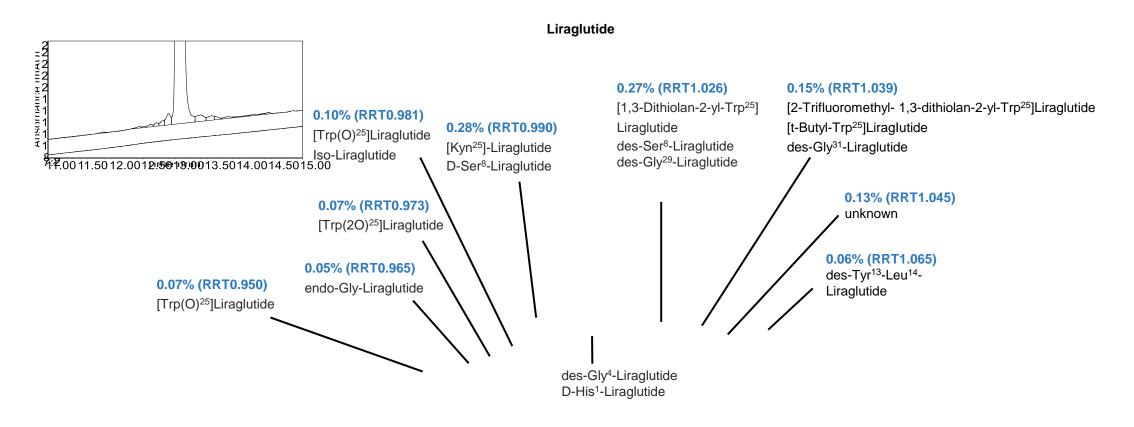
PREP LC-MS FATE & PURGE HIGH EFFICIENCY

- Impurities identified by MS and spiking experiments in case of isomers
- Define critical impurities for fate & purge
- The data set allows definition of regions where purge factors of 100% are achieved
- No synthesis and fate and purge study will be necessary for unambiguously assignable substances





LIRAGLUTIDE PURITY METHOD



Reporting threshold: 0.05 %



LIRAGLUTIDE BATCH COMPARISON

Batch	Batc	h 1	Bat	ch 2	Bat	ch 3	Bat	ch 4	Bat	ch 5	Bat	ch 6	Bat	ch 7	Bat	ch 8
Assignment	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm	% Area ⁽¹⁾	Δm
RRT 0.950	0.07%	+16	<	-	0.06%	+16	<	-	<	-	<	-	<	-	<	-
RRT 0.96	<	-	<	-	0.05%	+57	0.05%	+57	<	-	<	-	<	-	<	-
RRT 0.973	0.07%	+32	0.07%	+32	0.06%	+32	0.06%	+32	<	-	0.06%	+32	<	-	<	-
RRT 0.981	0.07%	+16	0.07%	+16	0.09%	+16	0.09%	+16	0.09%	+16	0.08%	+16	0.05%	+16	0.05%	+16
Impurity 5								±0		±0		±0				
RRT 0.990	0.20%	+4	0.17%	+4	0.18%	+4	0.23%	+4	0.25%	+4	0.11%	+4	0.09%	+4	0.08%	+4
Impurity 1								±0		±0		±0		±0		±0
RRT 1.026	0.27%	+104	0.20%	+104 (1)	0.19%	+104 (6)	0.23%	-87 (1)	0.11%	-87 (2)	0.19%	-87 (4)	0.11%	-87	0.11%	-87 (4)
				-87 (1)		-87 (1)		-57 (1)		-57 (1)		-57 (1)				-57 (1)
Impurity 2						-57 (1)										
RRT 1.039	0.14%	+172	0.12%	+172 (2)	0.10%	+172	0.12%	+172 (5)	0.15%	+172 (2)	0.08%	+172 (1)	0.05%	+172 (1)	0.07%	+172 (1)
Impurity 3				+56 (1)				-57 (4)		-57 (1)		-57 (1)		-57 (1)		-57 (1)
										+56 (1)				+56 (1)		+56 (1)
RRT 1.045	0.13%	+115	0.08%	+115	0.09%	+115	0.05%	n.d.	0.13%	n.d.	<	-	<	-	<	-
Impurity 4																
RRT1 .060/1 065	0.06%	-276	<	-	<	-	0.06%	-276	0.06%	-276	<	-	<	-	<	-
RRT1.110	<	-	<	-	<	-	<	-	<	-	<	-	0.07%	+14	0.05%	+14
RRT 1.234	0.06%	+3632	<	-	<	-	<	-	<	-	<	-	<	-	<	-
Purity ⁽¹⁾	98.9	9%	99.	.3%	99.	.2%	99	.1%	99.	.1%	99	.4%	99.	.8%	99.	.7%

(1) Values taken from release analysis



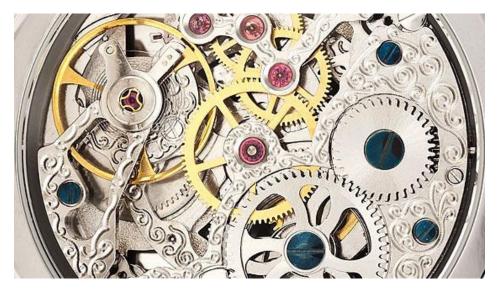
LIRAGLUTIDE IMPURITY SPECIFICATIONS

RRT	Δm [u]	Proposed Structure	max. %area (UV) found	Present in RLD	Impurity specified
0.981	+16	[Trp(O) ²⁵]Liraglutide	0.10%	Yes	Specified impurity 5
0.990	+4	[Kyn ²⁵]Liraglutide	0.25%	No	Specified impurity 1
0.990	±0	iso-Liraglutide, D-Ser8	0.23%	Yes	Specified impurity 1
	+104	2-(1,3-Dithiolan-2-yl-Trp ²⁵]Liraglutide		No	
1.021 / 1.026	-87	des-Ser ⁸ -Liraglutide	0.27%	No	Specified impurity 2
	-57	des-Gly ²⁹ -Liraglutide		No	
	+172	[2-Trifluoromethyl-1,3-dithiolan-2-yl- Trp ²⁵]Liraglutide		No	
1.039	+56	[t-Butyl-Trp ²⁵]Liraglutide	0.15%	No	Specified impurity 3
	-57	des-Gly ³¹ -Liraglutide		Yes	
1.045	+115	unknown	0.13%	No	Impurity not present with final process



RELEASE METHOD AGILITY AND EXPERTISE IS CRUCIAL FOR THE SUCCESS OF COMPLEX PEPTIDE THERAPEUTICS

- Complex Peptide NCEs require product-specific method development
- Quality control strategy must fulfill the needs of various stakeholders
- Complex impurity profiles need to be addressed with tailored analytics
- Tight ANDA-driven requirements can be met for large synthetic peptides





Cutting-edge analytical capabilities ensure development success and patient safety



GREAT TEAM KEY TO SUCCESS

Analytical Development

Agron Selami, Michael Naeff, Patrik Plattner, Constanze Schmies, Jürgen Opitz

CMC / Projekt Management:

Michael Berger, Michael Wollmann

Review/Mentoring:

Roland Eberli











THANK YOU



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UNDERSTANDING AGGREGATION BEHAVIOUR IMPORTANT FOR PROCESS DEVELOPMENT AND QC

QC Testing

HPLC SEC

- Determination of HMWP
- Quantitative, robust, sensitive (LOQ 0.1%)
- Routinely applicable for release
- MS suitable

Aggregation Screening

DLS

Measure kintetics to detect molecules with larger hydrodynamic radius

THT Assay

Linear fluorescence assay relating to the original amount of seeds

CD Spectroscopy

Detect secondary structures Comparative measurement API formulation / RLD

1nm	10 nm	100 nm	1 µm	10 µm	100 µm	1 mm	1 cm		
Monomer	Oligomer	Higher Order	Soluble Aggregates	Insoluble Aggregates					
		Subvisible	Subvisible			Visible			

